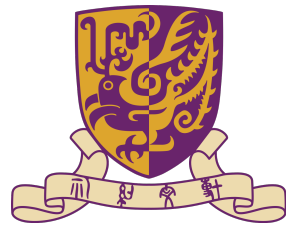




香港中文大學醫學院
Faculty of Medicine
The Chinese University of Hong Kong



Small molecules targeting RNA for novel antibiotic discovery

Name: Tsz Fung Tsang, year 3 PhD

Supervisor: Dr. Xiao Yang

Date : 6 December 2022

Overview

The need of discovering new drugs

Bacterial non-coding RNA as the new drug target

The potential druggability of bacterial riboswitch

New drugs for global public health issues

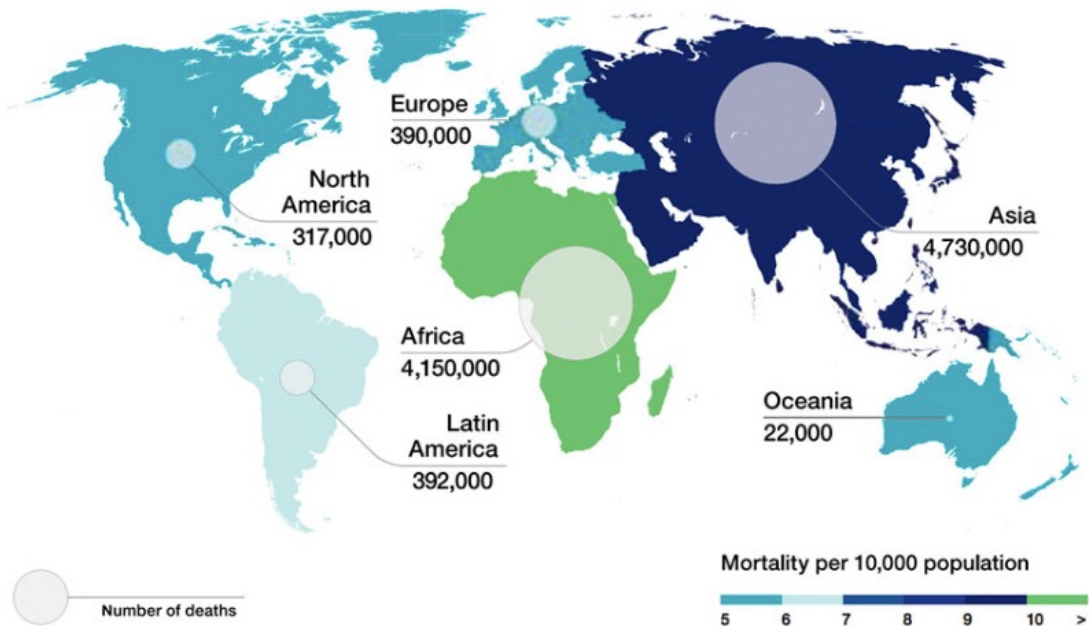
New Disease e.g. COVID-19

Cumulative deaths: 6 602 552

Data retrieved 23 Nov 2022
(WHO, 2020)

Antimicrobial resistance

Deaths attributable to AMR every year by 2050



(O' Neill, 2014)

For currently untreatable disorder

Spinal muscular atrophy (SMA) is characterized by progressive muscular atrophy and weakness.

Analysis of the long-term efficacy of nusinersen on SMA is lacking.

Homozygous deletions or mutations in the **survival motor neuron 1 (SMN1)** gene cause decreased functional **SMN protein**.

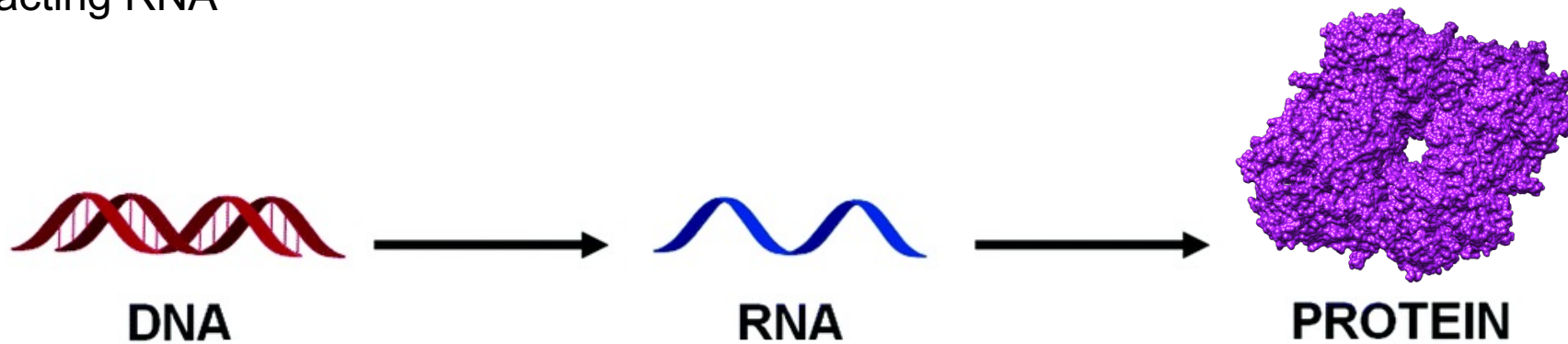
Nusinersen has been shown to effectively increase full-length SMN protein levels.

The infographic features a silhouette of a person in a wheelchair, a diagram of a neuron, and a 3D model of the SMN protein. An arrow labeled 'SMN' points down from the neuron diagram to the protein model, and another arrow labeled 'SMN' points up from the protein model to a bottle of Nusinersen.

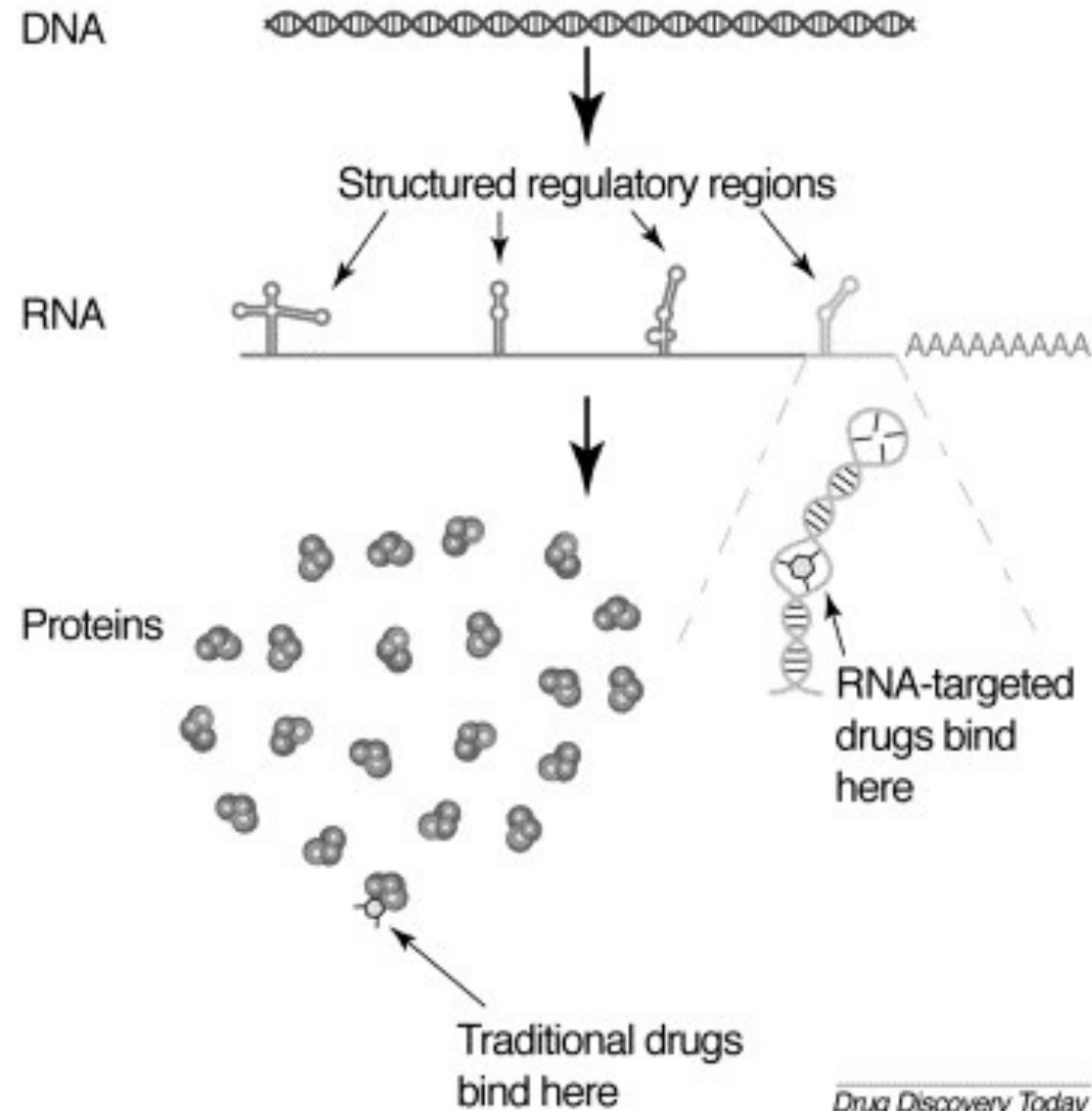
(American Academy of Neurology, 2019)

New potential drug targets from the discovery of non-coding RNA

lnc RNA	Long non-coding RNA
snoRNA	Small nucleolar RNA
miRNA	microRNA
snRNA	Small nuclear RNA
siRNA	Small interfering RNA
piRNA	Piwi-interacting RNA

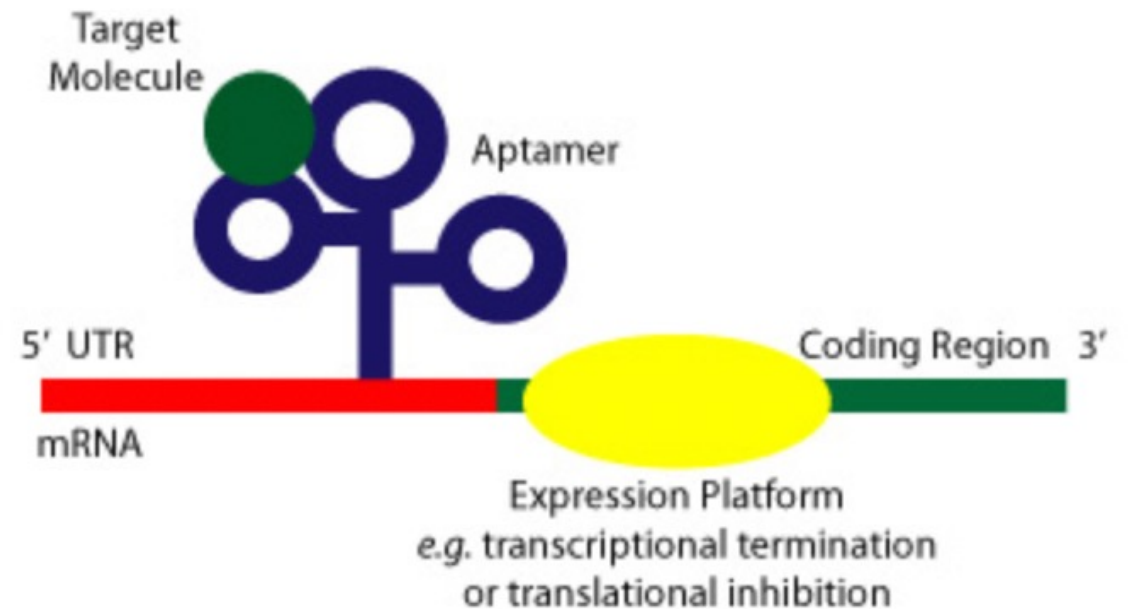


Bacterial non-coding RNA for drug targeting



Riboswitch

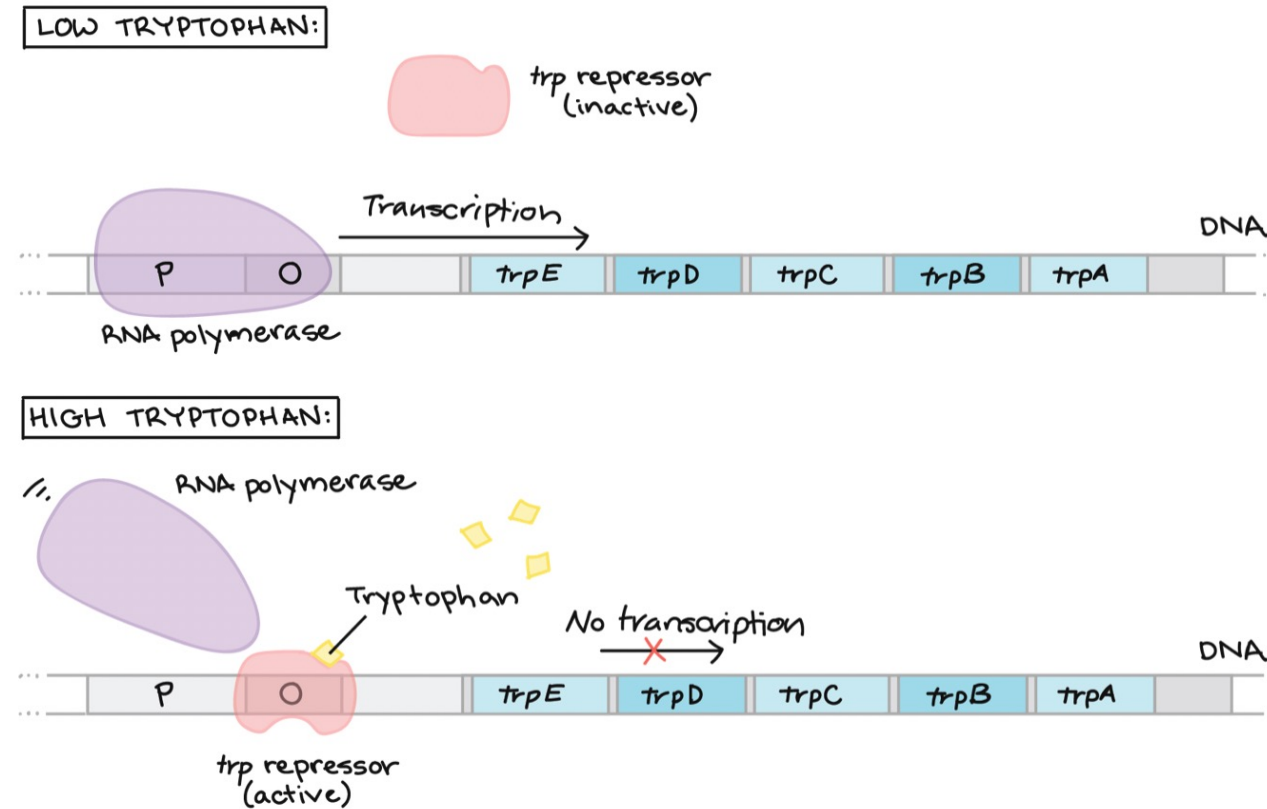
The element at 5' untranslated region (UTR) of mRNA



Bacterial non-coding RNA regulating gene expression

Protein-mediated feedback control

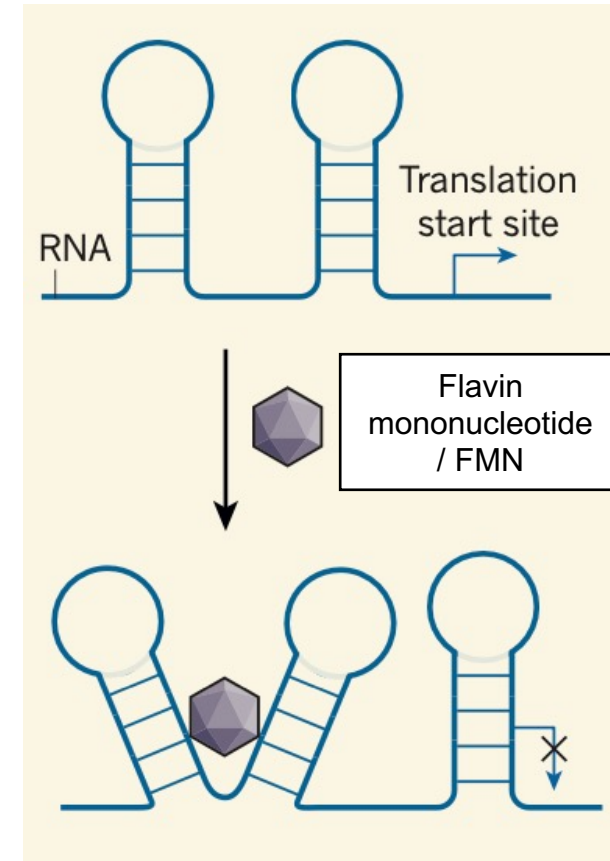
Example: Tryptophan (trp) biosynthesis



The excess tryptophan activates the transcription repressor (Babitzke and Gollnick., 2001).

RNA-mediated feedback control

Example: Riboflavin / vitamin B2 biosynthesis



FMN binds to the RNA aptamer for transcription inhibition (Hermann., 2015).

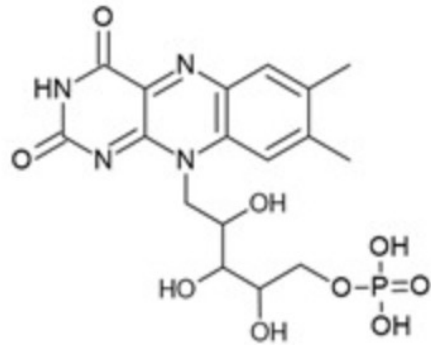
Riboswitches discovered in human pathogens

1 to 4% of all bacterial genes are regulated by riboswitches.

More than 35 different classes of riboswitches for a diverse set of metabolites have been identified.

Riboswitch	Type	Cognate Ligand
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FMN	Off	
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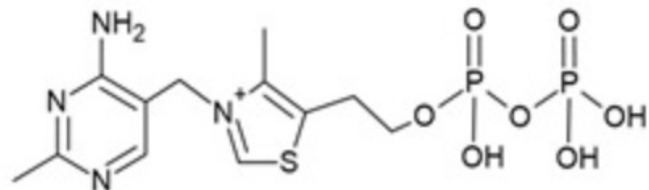


flavin mononucleotide (FMN)

FMN riboswitch identified in 41 pathogens

Acinetobacter baumannii, *Pseudomonas aeruginosa*,
Staphylococcus aureus, and *Streptococcus pneumoniae*

TPP	Off	
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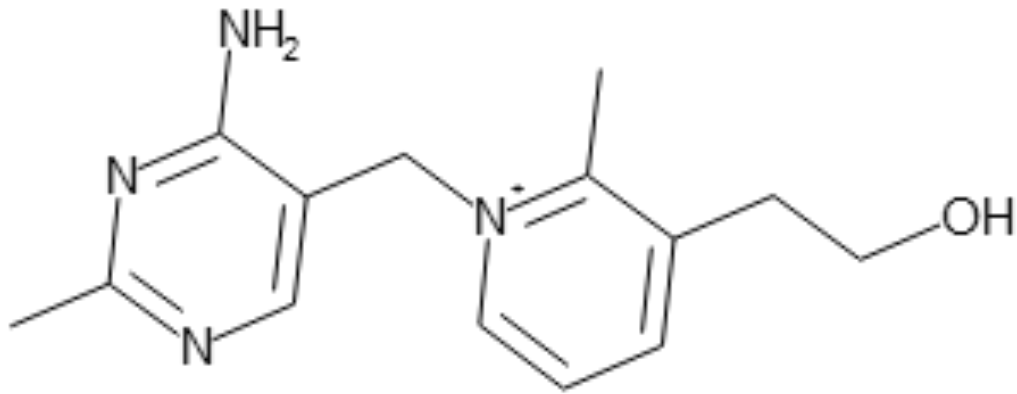


thiamine pyrophosphate (TPP)

TPP riboswitch identified in 48 pathogens: e.g.

A. baumannii, *P. aeruginosa*, *Mycobacterium tuberculosis*

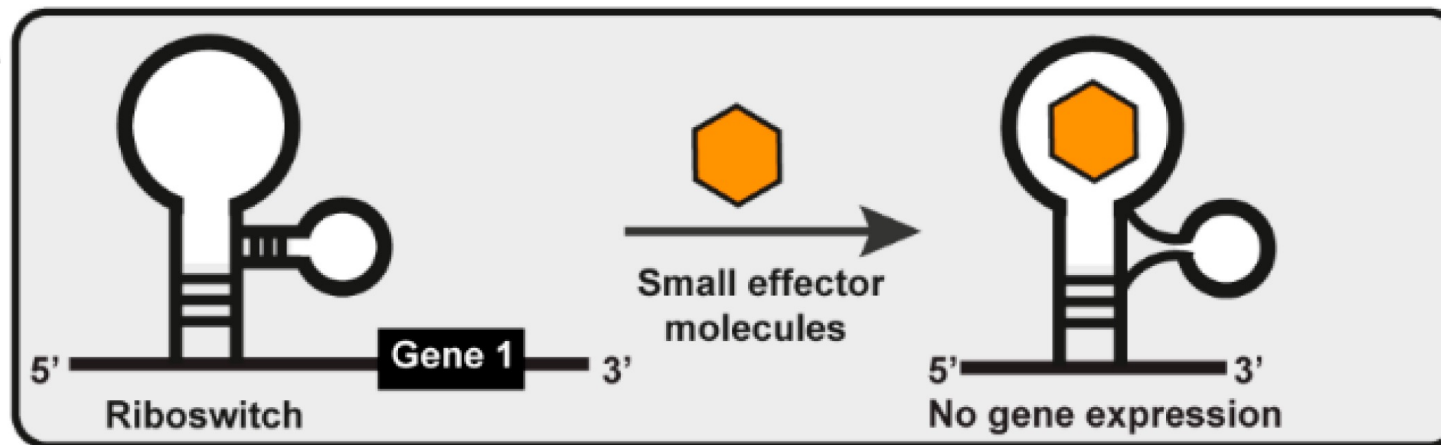
Disrupting bacterial riboswitch function for antibacterial activity



Pyrithiamine

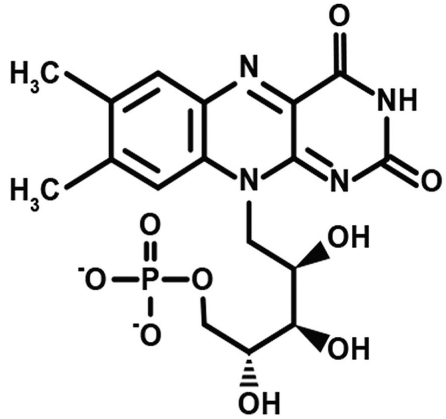
Pyrithiamine is an antibiotic discovered in 1943 (Woodley and White., 1943).

Recently proven to target bacterial TPP riboswitch (Sudarsan *et al.*, 2005).

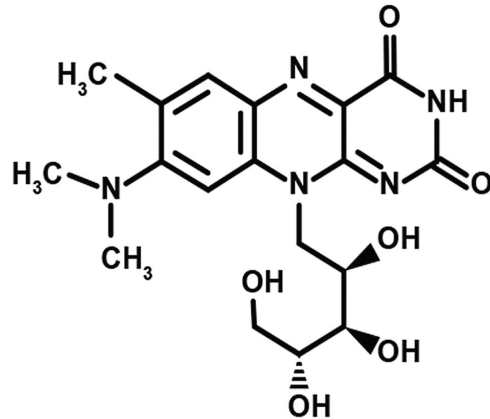


Proposed mode of action of inhibitors

Targeting FMN riboswitch by roseoflavin



FMN



Roseoflavin

Roseoflavin is a nature pigment produced in *Streptomyces davawensis*

Minimum inhibitory concentration (MIC)

MRSA RN4220: 0.06 µg/mL

MRSA COL: 0.06 µg/mL

Futher characterisation:

- Roseoflavin showed reduced bacterial burden in the murine infection model.
- Lethargy and ruffled fur in mice were observed.

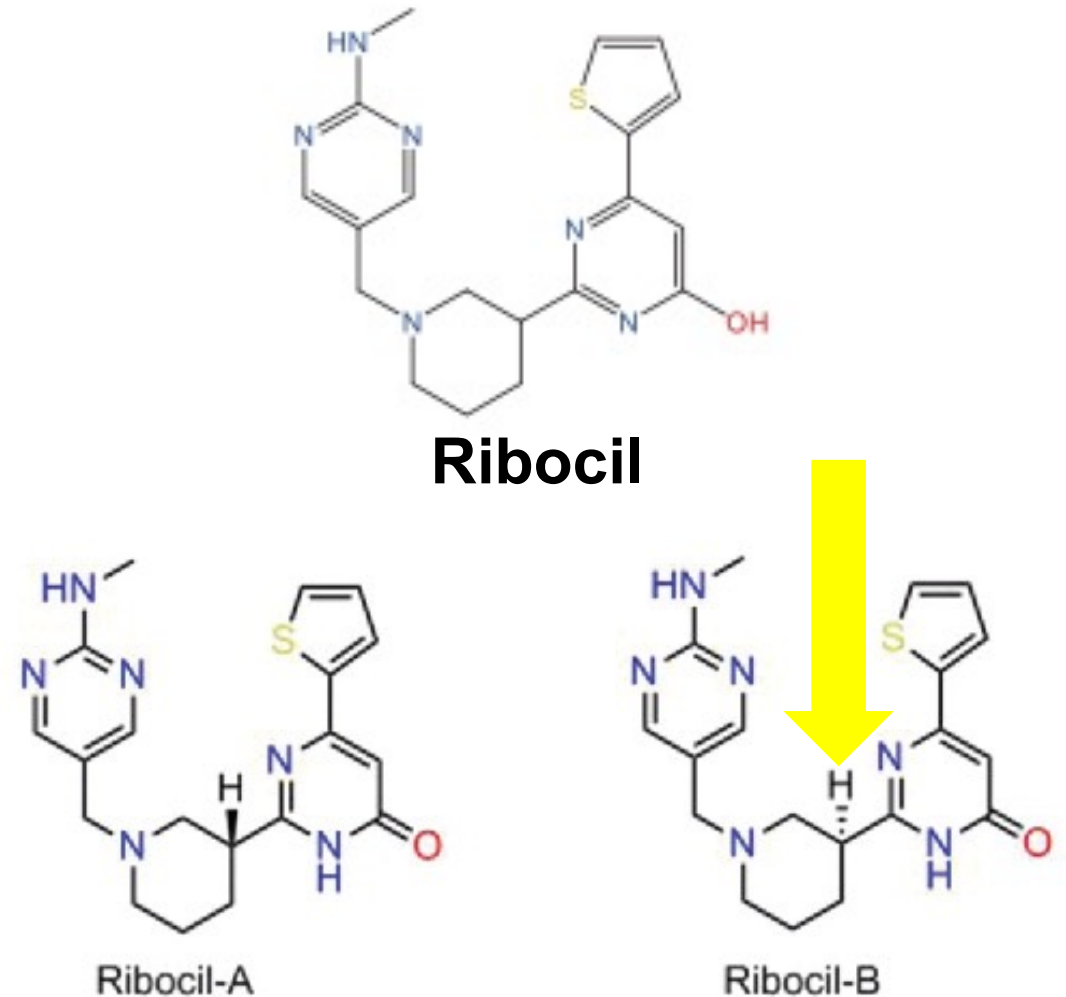
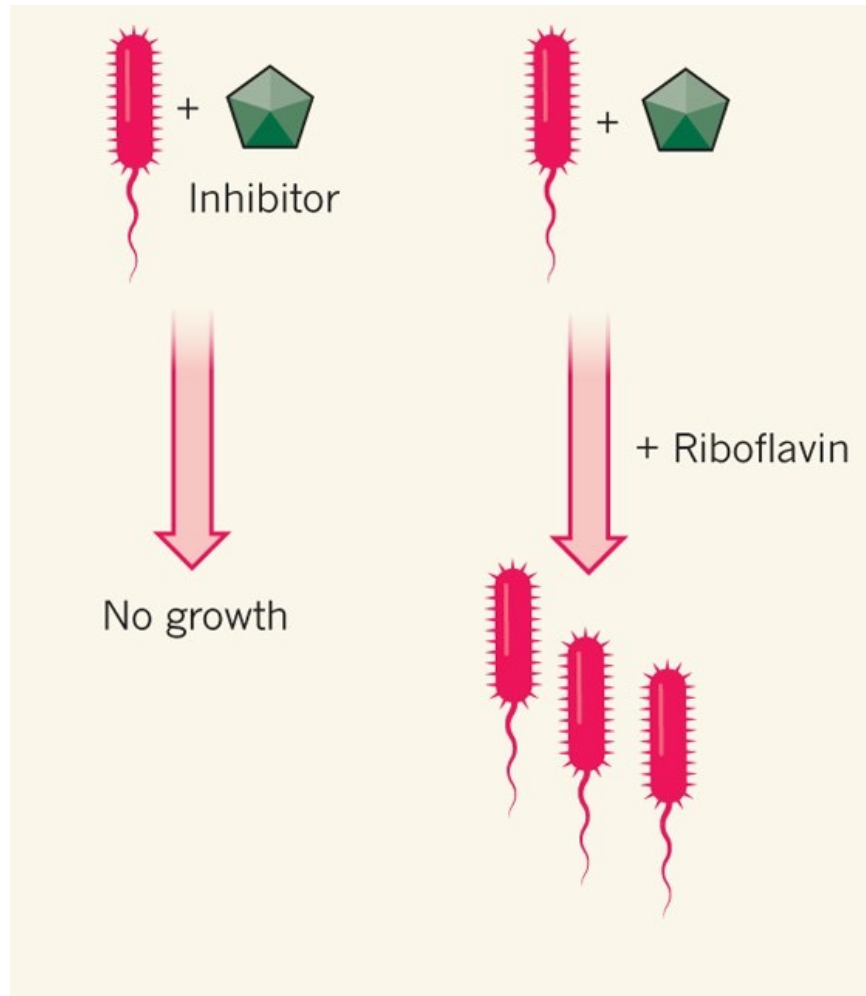
Concerns on the use of roseoflavin

1. Off-target effects in the murine infection model.
2. Structural and functional conservation between bacterial and human flavoenzymes.

Possible solution:

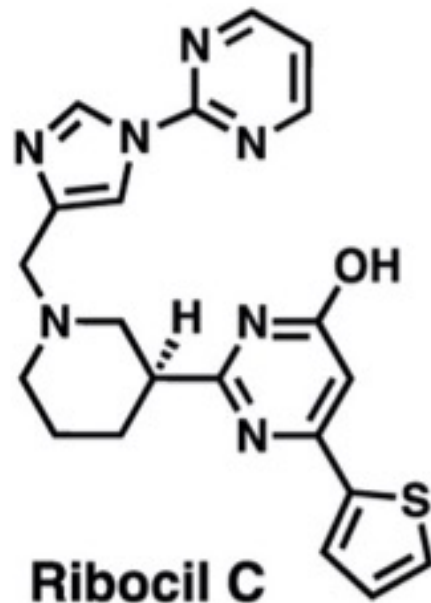
Search for a ligand that is not structurally similar to FMN.

Inhibitor screening for different compounds



Screened by culturing *E. coli* and compounds with / without riboflavin supplementation.

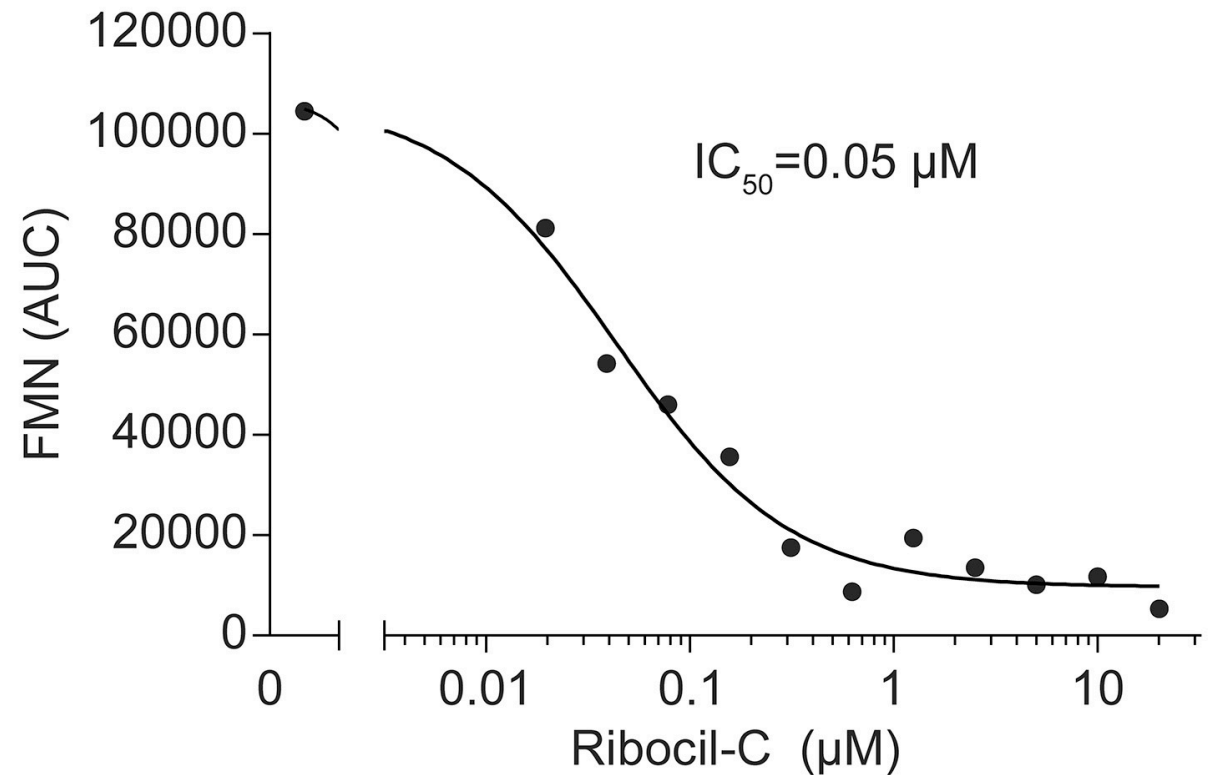
Ribocil C showing antimicrobial effect on *E. coli*



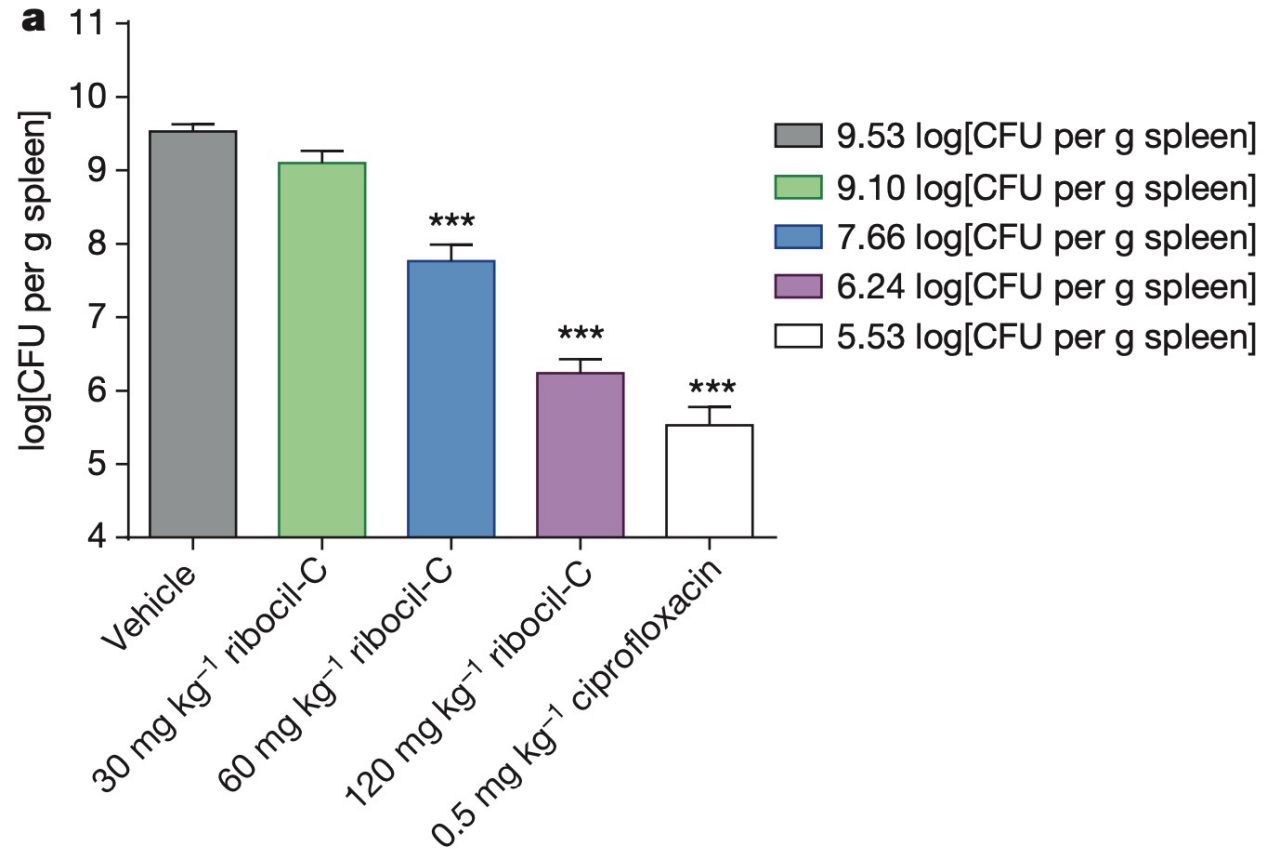
MIC against *E. coli*.

MB5746: 2 $\mu\text{g/mL}$

$\Delta\text{toI/C}$ strain: 0.25 $\mu\text{g/mL}$



Ribocil C with antibacterial effect *in vivo*



Bacterial burden (CFU per g spleen)

Mice infected with *E. coli* MB5746

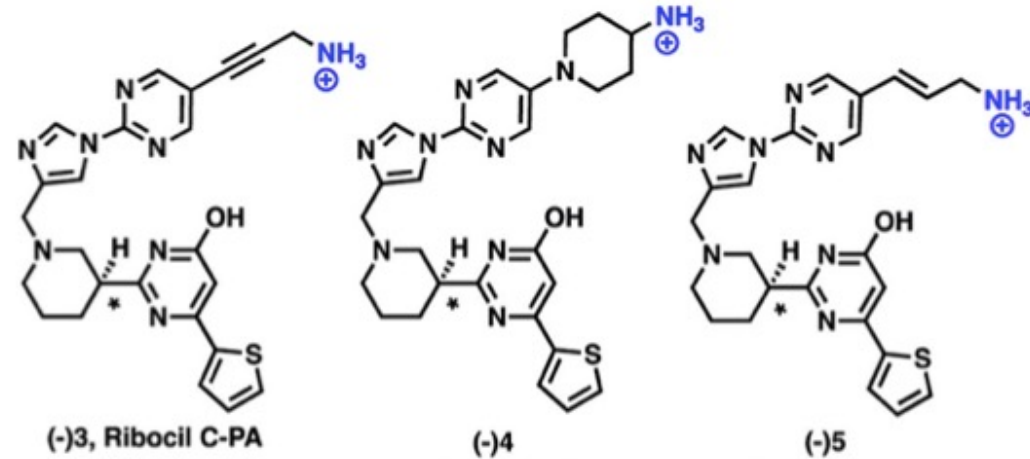
Infection: Intraperitoneal injection with *E. coli* MB5746.

Treatment: Subcutaneous injection with ribocil-C or ciprofloxacin at 30 / 60 / 120 mg/kg.

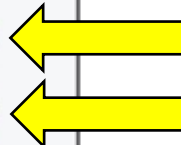
Finding: Reduction in bacterial burden when ribocil C was administered.

Limitation of ribocil C: No activity against wild type Gram-negative pathogens.

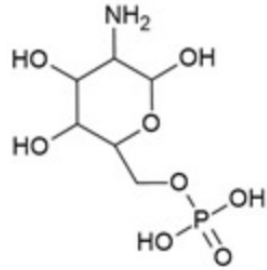
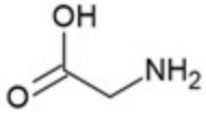
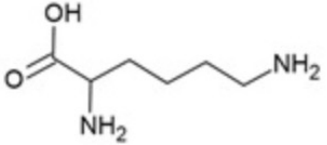
Ribocil C derivatives with improved antimicrobial activities against Gram-negative pathogens



Compound MIC ($\mu\text{g/mL}$)		(-)3	(-)4	(-)5
MC <i>E. coli</i>	<i>E. coli</i> ΔrfaC JW3596	0.5	4	1
	<i>E. coli</i> ΔtolC JW5503	0.25	0.5	0.25
GN Pathogens	<i>E. coli</i> BW25113	4	8	8
	<i>E. cloacae</i> ATCC BAA-2341	4	8	4
	<i>K. pneumoniae</i> ATCC 27736	4	8	4
	<i>A. baumannii</i> ATCC 2093	64	>64	64
	<i>P. aeruginosa</i> PAO1	>64	>64	>64



Other potentially druggable riboswitches

Riboswitch	Type	Cognate Ligand	
Fluoride	On	F ⁻	<i>A. baumannii</i> , <i>P. aeruginosa</i> , <i>E. faecium</i>
<i>glmS</i>	Off	 glucosamine-6-phosphate (GlcN6P)	<i>S. aureus</i> , <i>E. faecium</i>
Glycine	On	 glycine	<i>S. pneumoniae</i> , <i>Neisseria gonorrhoeae</i> , <i>S. aureus</i> , <i>A. baumannii</i> , <i>M. tuberculosis</i>
Lysine	Off	 lysine	<i>E. faecium</i> , <i>S. aureus</i>

Conclusion

- RNA can function more than an intermediate of DNA and protein.
- Bacterial non-coding RNA like riboswitch maybe the potential target for developing new antibiotics.
- Limitation: There are not enough knowledge about riboswitch.



Thanks for listening

Q&A

References

- Babitzke, P., & Gollnick, P. (2001). Posttranscription initiation control of tryptophan metabolism in *Bacillus subtilis* by the trp RNA-binding attenuation protein (TRAP), anti-TRAP, and RNA structure. *Journal of bacteriology*, 183(20), 5795–5802. <https://doi.org/10.1128/JB.183.20.5795-5802.2001>
- Bose, D. P. (2022, June 27). *From DNA to protein; the central dogma of molecular biology*. News-Medical.net. Retrieved November 24, 2022, from <https://www.azolifesciences.com/article/From-DNA-to-Protein3b-The-Central-Dogma-of-Molecular-Biology.aspx>
- https://tmedweb.tulane.edu/pharmwiki/doku.php/ribosomal_antibiotics
- Blount, K. F., & Breaker, R. R. (2006). Riboswitches as antibacterial drug targets. *Nature biotechnology*, 24(12), 1558–1564. <https://doi.org/10.1038/nbt1268>
- Childs-Disney, J. L., Yang, X., Gibaut, Q. M. R., Tong, Y., Batey, R. T., & Disney, M. D. (2022). Targeting RNA structures with small molecules. *Nature reviews. Drug discovery*, 21(10), 736–762. <https://doi.org/10.1038/s41573-022-00521-4>
- Costales, M. G., Childs-Disney, J. L., Haniff, H. S., & Disney, M. D. (2020). How We Think about Targeting RNA with Small Molecules. *Journal of medicinal chemistry*, 63(17), 8880–8900. <https://doi.org/10.1021/acs.jmedchem.9b01927>
- Ecker, D. J., & Griffey, R. H. (1999). RNA as a small-molecule drug target: doubling the value of genomics. *Drug discovery today*, 4(9), 420–429. [https://doi.org/10.1016/s1359-6446\(99\)01389-6](https://doi.org/10.1016/s1359-6446(99)01389-6)
- Howe, J. A., Wang, H., Fischmann, T. O., Balibar, C. J., Xiao, L., Galgoci, A. M., Malinverni, J. C., Mayhood, T., Villafania, A., Nahvi, A., Murgolo, N., Barbieri, C. M., Mann, P. A., Carr, D., Xia, E., Zuck, P., Riley, D., Painter, R. E., Walker, S. S., Sherborne, B., ... Roemer, T. (2015). Selective small-molecule inhibition of an RNA structural element. *Nature*, 526(7575), 672–677. <https://doi.org/10.1038/nature15542>
- Khan Academy. (n.d.). *The TRP operon (article)*. Khan Academy. Retrieved November 30, 2022, from <https://www.khanacademy.org/science/ap-biology/gene-expression-and-regulation/regulation-of-gene-expression-and-cell-specialization/a/the-trp-operon>
- Kavita, K., & Breaker, R. R. (2022). Discovering riboswitches: the past and the future. *Trends in biochemical sciences*, S0968-0004(22)00234-1. Advance online publication. <https://doi.org/10.1016/j.tibs.2022.08.009>
- Lee, E. R., Blount, K. F., & Breaker, R. R. (2009). Roseoflavin is a natural antibacterial compound that binds to FMN riboswitches and regulates gene expression. *RNA biology*, 6(2), 187–194. <https://doi.org/10.4161/rna.6.2.7727>
- Meyer, S. M., Williams, C. C., Akahori, Y., Tanaka, T., Aikawa, H., Tong, Y., Childs-Disney, J. L., & Disney, M. D. (2020). Small molecule recognition of disease-relevant RNA structures. *Chemical Society reviews*, 49(19), 7167–7199. <https://doi.org/10.1039/d0cs00560f>
- Mironov, A. S., Gusarov, I., Rafikov, R., Lopez, L. E., Shatalin, K., Kreneva, R. A., Perumov, D. A., & Nudler, E. (2002). Sensing small molecules by nascent RNA: a mechanism to control transcription in bacteria. *Cell*, 111(5), 747–756. [https://doi.org/10.1016/s0092-8674\(02\)01134-0](https://doi.org/10.1016/s0092-8674(02)01134-0)
- Nusinersen super effective in later-onset SMA!* TreatSMA. (2019, September 22). Retrieved November 24, 2022, from <https://www.treatsma.uk/2019/05/nusinersen-super-effective-in-later-onset-sma/>
- O' Neill, J. Antimicrobial resistance: tackling a crisis for the health and wealth of nations. The review on antimicrobial resistance, 2014, 4-16
- Panchal, V., & Brenk, R. (2021). Riboswitches as Drug Targets for Antibiotics. *Antibiotics (Basel, Switzerland)*, 10(1), 45. <https://doi.org/10.3390/antibiotics10010045>
- Protein synthesis inhibitors. ribosomal_antibiotics* [TUSOM | Pharmwiki]. (n.d.). Retrieved November 24, 2022, from https://tmedweb.tulane.edu/pharmwiki/doku.php/ribosomal_antibiotics
- Wang, H., Mann, P. A., Xiao, L., Gill, C., Galgoci, A. M., Howe, J. A., Villafania, A., Barbieri, C. M., Malinverni, J. C., Sher, X., Mayhood, T., McCurry, M. D., Murgolo, N., Flattery, A., Mack, M., & Roemer, T. (2017). Dual-Targeting Small-Molecule Inhibitors of the *Staphylococcus aureus* FMN Riboswitch Disrupt Riboflavin Homeostasis in an Infectious Setting. *Cell chemical biology*, 24(5), 576–588.e6. <https://doi.org/10.1016/j.chembiol.2017.03.014>
- Serganov, A., & Nudler, E. (2013). A decade of riboswitches. *Cell*, 152(1-2), 17–24. <https://doi.org/10.1016/j.cell.2012.12.024>
- Sudarsan, N., Cohen-Chalamish, S., Nakamura, S., Emilsson, G. M., & Breaker, R. R. (2005). Thiamine pyrophosphate riboswitches are targets for the antimicrobial compound pyrithiamine. *Chemistry & biology*, 12(12), 1325–1335. <https://doi.org/10.1016/j.chembiol.2005.10.007>
- Winkler, W., Nahvi, A., & Breaker, R. R. (2002). Thiamine derivatives bind messenger RNAs directly to regulate bacterial gene expression. *Nature*, 419(6910), 952–956. <https://doi.org/10.1038/nature01145>
- World Health Organization. (n.d.). *Who coronavirus (COVID-19) dashboard*. World Health Organization. Retrieved November 24, 2022, from <https://covid19.who.int/>